Novel deletion of SPAST in a Chinese family with hereditary spastic paraplegia

Yapei Feng1,*, MS, Xin Ke2,*, MD, Meng Zhai1, MS, Qian Xin1, MS, Yaoqin Gong1, PhD, Qiji Liu1, PhD

INTRODUCTION
Hereditary spastic paraplegia (HSP) belongs to a highly heterogeneous group of neurodegenerative disorders and is characterised by progressive spasticity and weakness of the lower limbs. The main pathological changes associated with the disorder are developmental failure or degeneration of motor axons in the corticospinal tract.1-2 Genetic studies have shown the extremely heterogeneous nature of HSP, and at least 46 genetic loci responsible for HSP have been identified in families with autosomal dominant (AD), autosomal recessive and X-linked inheritances, according to the HUGO Gene Nomenclature and OMIM (Online Mendelian Inheritance in Man) databases.3-5 HSPs can be classified into either pure or complicated forms, based on the clinical manifestations. The more common pure form of HSP usually presents four core features: (a) slowly progressive spastic gait; (b) increasing muscle tone in the lower limbs; (c) hyperreflexia; and (d) extensor plantar response.6-7 Complicated HSP manifests other neurological or extraneurological abnormalities, such as mental retardation, deafness, cerebellar ataxia, epilepsy, dysthria and ichthyosis. Although patients with pure HSP generally exhibit the disease’s main clinical features, considerable variation is observed in the age of the patient at disease onset, as well as in the rate of progression and severity of spasticity within and between families.1,2,5-8

The SPG4 gene, which has been mapped to chromosome 2p21-p22, is the most common causative gene of autosomal dominant hereditary spastic paraplegia (ADHSP). Spastic paraplegia type 4 (SPG4) accounts for approximately 40% of AD pedigrees and 6%–15% of sporadic cases.9 To date, more than 257 SPAST mutations have been reported,10 including missense, nonsense or splice site mutations, small insertions or deletions, and large duplications or deletions.11-14 SPG4 encodes spastin, a 616 amino acid protein, which is a member of the AAA cassette (adenosine triphosphatases associated with diverse cellular activities). Spastin is a microtubule-severing enzyme15 involved in remodelling protein complexes with adenosine triphosphate (ATP) hydrolysis and coordination of axonal microtubule interactions with the tubular endoplasmic reticulum network.16,17 In this context, we conducted a genetic study of a four-generation Chinese family with ADHSP, investigating the SPAST gene via linkage analysis and direct sequencing methodologies.

METHODS
The patients were from a Chinese family spanning four generations from Weihai, Shandong Province, China. HSP followed a pattern of AD inheritance in this family (Fig. 1). The genetic

1Key Laboratory for Experimental Teratology of the Ministry of Education and Department of Medical Genetics, Shandong University School of Medicine, Jinan, 2Department of Orthopaedics, Hospital of Weihai Economic and Technical Development Zone, Weihai, People’s Republic of China 3These two authors contributed equally as first author in this work.

Correspondence: Dr Qiji Liu, Professor, Key Laboratory for Experimental Teratology of the Ministry of Education and Department of Medical Genetics, Shandong University School of Medicine, Jinan 250012, People’s Republic of China. liuqiji@sdu.edu.cn
study, conducted on the members of this family, was approved by the ethics committee of the Shandong University School of Medicine. Signed, informed consent was obtained from all participants prior to the study. Based on Harding's criteria,\(^{15}\) an experienced neurologist performed neurological examinations on 11 family members and one of the spouses. The index patient (patient II10) underwent electromyography, magnetic resonance imaging, neurophysiological studies, cerebrospinal fluid examination and cognitive tests. Other related neural diseases were excluded. The clinical features of patients with this pedigree, who might represent individuals with a pure form of HSP, are presented in Table I. Blood samples were obtained from 11 family members (symptomatic n = 4; asymptomatic n = 7) and one spouse (unaffected). Leucocyte genomic DNA was extracted using standard techniques.

We first tested whether HSP in the family was caused by a mutation in any of the known HSP loci. As SPG4 is the most common causative gene for ADHSP and SPG4 mutation accounts for approximately 40% of AD pedigrees,\(^{36}\) we selected D2S2351 and D2S2347 (lying 0.2 Mb upstream and 0.7 Mb downstream of SPG4, respectively), two microsatellite markers with high heterozygosity in the population according to Marshfield genetic maps, for analysis.\(^{16}\) Polymerase chain reaction (PCR) amplification was performed in a 20 μl volume containing 8 μl of water, 4 μl of 5 × TNK50, 4 μl of 5 × deoxyribonucleotide triphosphates, 0.5 μl of each primer (10 μM), 2 μl of genomic DNA (40 ng/μl) and 1 μl of Taq DNA polymerase (1 U/μl; TianGen Biotech, Beijing, China), with an initial denaturation step of 5 min at 95°C to activate the polymerase, followed by 35 cycles of 95°C for 40 s, 55°C for 40 s, 72°C for 40 s and a final elongation at 72°C for 10 min. Predicted amplicon sizes were confirmed by agarose gel electrophoresis. The amplified fragments of D2S2351 and D2S2347 (patients II3, II5, II8, II10 and III11), with internal size standards, were 201–282 bp. Polymorphism was estimated using polyacrylamide gel electrophoresis (PAGE).

The primers used were designed to examine entire exonic sequences of SPAST (MIM 604277) via PCR amplification and direct sequencing. Large exons were divided into several overlapping PCR fragments. From the index patient’s sample, all exons, including exon-intron boundaries, were amplified by PCR and direct sequencing. The sequences of the other family members and 50 normal, unaffected individuals who were matched for geographical ancestry were confirmed by Sanger sequencing. Changes in exons were confirmed by repeat PCR and sequencing of both strands of the resulting product in ten family members other than the index patient and the spouse. The amplified fragments of exon 15, with internal size standards, were 373 bp with the use of the designed primers. Polymorphism in the amplified fragments of exon 15 of the SPAST gene was estimated in four panels by non-denatured PAGE (ND-PAGE) for the 11 family members. Amplified products with mutations were ligated to a pMD-18T vector and transformed into Escherichia coli (E. coli) bacteria. Detected changes in the exons were precisely defined by subcloning PCR and sequencing.

**RESULTS**

The phenotype of the four patients (patients III3, II8, II10 and III11) who were found to have HSP was late disease onset (age ≥ 45 years) and severe clinical manifestations. Three patients (patients III3, II10 and III11) had severe symptoms, while patient II8 showed severe symptoms that were relatively fewer in number. Symptoms in the proband (patient II10), a 69-year-old woman, began at the age of 45 years, and included progressively spastic gait, walking difficulty that required assistance, tendon

<table>
<thead>
<tr>
<th>Patient</th>
<th>Gender</th>
<th>Age at examination (yrs)</th>
<th>Age at onset (yrs)</th>
<th>Disability grade*</th>
<th>Ashworth scale*</th>
<th>Spastic gait</th>
<th>Hypereflexia in legs</th>
<th>Increased tone in legs</th>
<th>Extensor plantar reflex</th>
<th>Weakness in legs</th>
<th>Wasting in legs</th>
</tr>
</thead>
<tbody>
<tr>
<td>II1</td>
<td>M</td>
<td>–</td>
<td>45</td>
<td>4</td>
<td>4</td>
<td>+++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>II2</td>
<td>F</td>
<td>–</td>
<td>46</td>
<td>4</td>
<td>4</td>
<td>+++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>II3</td>
<td>F</td>
<td>69</td>
<td>45</td>
<td>4</td>
<td>4</td>
<td>+++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>II5</td>
<td>M</td>
<td>71</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>II8</td>
<td>F</td>
<td>65</td>
<td>45</td>
<td>3</td>
<td>3</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>II10</td>
<td>F</td>
<td>58</td>
<td>45</td>
<td>4</td>
<td>4</td>
<td>+++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>II11</td>
<td>M</td>
<td>61</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>III1</td>
<td>F</td>
<td>46</td>
<td>46</td>
<td>1</td>
<td>2</td>
<td>–</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>–</td>
</tr>
<tr>
<td>III4</td>
<td>F</td>
<td>45</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>III6</td>
<td>M</td>
<td>42</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>III10</td>
<td>F</td>
<td>38</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>III11</td>
<td>F</td>
<td>35</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>III12</td>
<td>F</td>
<td>36</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>IV1</td>
<td>M</td>
<td>23</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

*Disability grades: no mobility problems or slight stiffness of the legs (grade 1); moderate gait stiffness (grade 2); problems running but able to walk alone (grade 3); problems walking (grade 4); wheelchair-bound (grade 5). ‘Ashworth scale: no increase in muscle tone (scale 1); slight increase in tone giving a ‘catch’ when affected part is moved in flexion or extension (scale 2); more marked increase in tone, but affected part is easily flexed (scale 3); considerable increase in tone and difficult passive movement (scale 4); affected part is rigid in flexion or extension (scale 5).

= absent; +: mild; ++: moderate; +++: severe; F: female; M: male
hyperreflexia of the lower limbs and Babinski’s sign. Initial symptoms of walking difficulty and progressively spastic gait in four of our affected patients began at a similar age. The disease progressed slowly, and there was correlation of clinical severity with disease duration. None of the patients exhibited upper extremity features or urinary disturbances (urgency and/or incontinence). Disease onset in patient III1 began at the age of 46 years, with mild walking difficulty, increasingly reduced muscle tone in the lower limbs and hyperreflexia. Patients III6 and IV1 were asymptomatic and showed no abnormal features on neurological examination at the ages of 42 and 23 years, respectively. As most of the third/fourth generation family members had not yet reached the age of disease onset, we could not determine whether they were affected or not based on phenotype alone. The family members available for linkage analysis were thus fewer and we were unable to get a high logarithm of odds (LOD) score at D2S2347. We found that among the available family members, alleles of D2S2347 cosegregated with the phenotype. Although we could not confirm that SPG4 was the disease gene, we also could not exclude the possibility of it being the causative gene. There was no D2S2351 polymorphism found in the family.

Via Sanger sequencing, we were able to determine whether an SPG4 mutation was present in members of the Chinese family with ADHSP. Our results revealed a 14-bp heterozygous deletion (c.1630-1643delTACTCAGGAAGTGA) in exon 15 of SPAST, leading to a frameshift mutation and premature termination of translation (p.tyr544profsX28). The 14-bp heterozygous deletion was precisely defined by subcloning PCR products in E. coli and sequencing (Fig. 2). Our findings also identified cosegregation of the deletion and phenotype in other affected family members. This mutation was, however, found to be absent in 50 normal, unaffected individuals who were matched for geographical ancestry. Results of the ND-PAGE of the amplified fragments of exon 15 of SPAST showed that the affected family members (patients II3, II8, II10 and III1) carried a 14-bp deletion gene, as did two other patients who were asymptomatic (patients III6 and IV1), but those of the 50 normal, unaffected individuals who were matched for geographical ancestry were homozygous with no mutation.

DISCUSSION

In this study, combined linkage analysis and sequencing enabled the detection of a novel mutation in the SPAST gene that was causing ADHSP in a four-generation Chinese family. Previous reports have shown that HSP, which has a strong genetic basis, is clinically heterogeneous. The age of disease onset ranges from 1–77 years and the severity of symptoms differs greatly in many families with ADHSP. In the four-generation Chinese family with ADHSP studied here, the phenotypes of the four affected patients were late disease onset (≥ 45 years) and severe clinical manifestations (with no upper extremity features or urinary disturbances – urgency and/or incontinence), although bladder disturbances were previously reported in approximately 50% of affected individuals.19,20

Most of the SPAST mutations reported include missense, nonsense or splice site mutations, small insertions or deletions, and large duplications and deletions. In our study, we detected a novel 14-bp heterozygous deletion (c.1630-1643delTACTCAGGAAGTGA) in exon 15 of the SPAST gene, which led to a frameshift mutation with a premature stop codon. However, this mutation was found to be absent in 50 normal individuals who were matched for geographical ancestry. Sequence analysis revealed the presence of the 14-bp deletion...
in two asymptomatic patients (patients III6 and IV1) who showed no abnormal features on neurological examination at age 42 years and 23 years, respectively. We could not determine whether incomplete dominance existed in this four-generation Chinese family with ADHSP, as the phenomenon’s penetrance might be age-dependent or incomplete, or manifest subtle symptoms. Subtle symptoms may progress slowly and go unnoticed for years. The variation seen in exon 15 was at a highly conserved position, which predicted functional impact and cosegregation with the phenotype. Mutation (p.tyr544profsX28) between amino acids 544 and 571, with a premature termination, is predicted to disrupt the highly conserved functional domain (the AAA cassette), which includes the ATP binding and hydrolysis sites. The likely pathogenic mechanism for the frameshift mutation in our study is haploinsufficiency, which leads to unstable mRNA levels and absence or low level of expression of proteins. Although haploinsufficiency (loss-of-function mechanism) is currently the most accepted view, gain-of-function of a ‘toxic’ or dominant negative activity could also partly explain this pathogenic mechanism of SPAST mutation.

Hence, the mechanism of HSP (via mutation in SPG4) is still a subject of much dispute. In conclusion, we investigated the SPAST gene via linkage analysis and sequencing in a four-generation Chinese family who exhibited ADHSP. We found a novel mutation in exon 15 of the SPG4 gene in members of this family, which was absent in unaffected individuals who were matched for geographical ancestry. The mutation in a highly conserved AAA cassette spanning exon 7 to 16 was predicted to have functional impact and found to completely cosegregate with the HSP phenotype. Our result expands the mutation spectrum of SPG4. The identification of this novel deletion in SPAST provides an opportunity to further study and increase our understanding of the mechanism of axonal neurodegeneration in HSP, which may have important therapeutic implications. As better understanding of the function of spasticity is important, further phenotype-genotype correlation studies will be required to help explain the variable expression of the SPG4 form for the prevention, diagnosis, and treatment of HSP.

ACKNOWLEDGEMENTS

We thank the patients and their families for their participation. This work was supported by grants from the National Natural Science Foundation of China (grant no. 81072452) and the Research Award Fund for Outstanding Young Scientist of Shandong Province (2007BS0327).

REFERENCES